

THREE-DIMENSIONAL MULTI-COLOUR SINGLE MOLECULE LOCALISATION MICROSCOPY AT CRYOGENIC TEMPERATURES

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Cryo-vitrification is an established technique allowing the preservation of the ultrastructural morphology in biological specimen and is superior to artefact-inducing chemical fixation commonly used in fluorescent sample preparation at room temperatures [1]. Cryogenic temperatures also reduce the photodecomposition rate of fluorescent molecules leading to an improved localisation precision and hence resolution in single-molecule localisation microscopy (SMLM) [2]. The advantages outlined would only be achievable through the use of high-numerical aperture (typically $NA > 1.4$) oil immersion objective lenses which are not compatible at cryogenic temperature due to freezing. In a recent paper [3] we describe the use of a solid immersion lens (*superSIL*) coupled to a dry objective (0.55 NA) to effortlessly achieve an effective $NA = 2.17$ at cryogenic temperatures. This enables us to routinely acquire SMLM images with localisation precision of ~ 8 nm and lateral resolution of < 20 nm on plunge-frozen bacterial and yeast cells in liquid nitrogen vapour. We have subsequently extended this technique to enable simultaneous two-colour SMLM imaging of multiple proteins in cells. Furthermore, we have modelled and applied Point Spread Function (PSF) engineering to the *superSIL* microscope to characterise axial distance in the ~ 250 nm depth of field.

We believe this method paves the way for significant improvements to cryo-correlative light and electron microscopy (cryo-CLEM) research and associated applications.

References

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