

# LIGHT SHEET FLUORESCENCE MICROSCOPY IMAGING FOR THE DEVELOPMENT OF AN *IN VITRO* TISSUE BOUNDARY 3D TUMOUR MODEL USING SELF-ASSEMBLED HYDROGEL

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Multicellular Tumour spheroids (MCTS) have become increasingly popular and widely regarded as an excellent 3D *in vitro* model to examine tumour drug penetration [1,2] and the influence of tumour microenvironment [3]. Yet, imaging MCTS has been challenging using conventional confocal or multiphoton microscopy, due to their limited depth, working distance and phototoxicity as well as the specimen preparation.

Light sheet fluorescence microscopy (LSFM) overcomes many of these issues [4] and here we describe the specimen preparation of spheroids for 4D imaging via Zeiss Lightsheet Z.1 and the Leica TCS SP8 DLS. We have used these preparations to examine SK-N-BE(2) neuroblastoma spheroids “sandwiched” between two hydrogels (asymmetric model).

Our aim is to mimic the growth of a tumour on the boundaries between two tissue types with extracellular matrix (ECM) of different mechanical properties and morphologies.

Self-assembled peptide hydrogels are a subclass of biomimetic materials able to mimic the nanostructure of the ECM [5] in biomedical applications. They are often easily functionalized, stable in common cell culture media and optically transparent. Moreover, the versatility of these biomimetic materials allows us to display a wide range of physiological-like mechanical properties characterized by Atomic Force Microscopy Nano indentation.

The use of tunable hydrogel scaffolds as ECM mimics for MCTS LFSM 4D imaging provides an adaptable mechanotransduction platform for an *in vitro* tissue boundaries tumour model.

[1] J.S. Basuki; H.T.T. Duong; A. Macmillan; R.B. Erlich; L. Esser; M.C. Akerfeldt; R.M. Whan; M. Kavallaris; C. Boyer; T.P. Davis, “Using fluorescence lifetime imaging microscopy to monitor theranostic nanoparticle uptake and intracellular doxorubicin release”, *ACS nano*, **7** (11), 10175-10189 (2013).

[2] S.M. Sagnella; H. Duong; A. macmillan; C. Boyer; R. Whan; J.A. mccarroll, T.P. Davis, M. Kavallaris, “Dextran-based doxorubicin nanocarriers with improved tumor penetration,” *Biomacromolecules*, **15** (1), 262-275 (2013).

[3] J. Wojciechowski; A. Martin; A. Mason; C. Fife; S. Sagnella; M. Kavallaris; P. Thordarson, “Choice of Capping Group in Tripeptide Hydrogels Influences Viability in the Three-Dimensional Cell Culture of Tumor Spheroids,” *ChemPlusChem* (2016) DOI: 10.1002/cplu.201600464.

[4] A.A. Yakimovich and V. Andriasyan, “Fast Imaging of Cellular Spheroids with Light Sheet Fluorescence Microscopy,” *Zeiss Application Note* (2013).

[5] E.Y. Du; A.D Martin; C. Heu; P. Thordarson, “The Use of Hydrogels as Biomimetic Materials for 3D Cell Cultures,” *Australian Journal of Chemistry* **70**, 1-8 (2016).